

DNA Content Protocol

Reagents

Propidium Iodide (PI) 1mg/mL stock:	[PI]_F = 40ug/mL	PI Master Mix
RNase (DNase-free) 10mg/mL stock:	[RNase]_F = 100ug/mL	PI 40uL
PBS, Ca ⁺⁺ , Mg ⁺⁺ - free		RNase 10uL
70% Ethanol		PBS <u>950uL</u>
		1,000uL

Procedure

1. Wash 1-2X10⁶ single cells (perform an initial cell count with hemocytometer) in cold PBS.
2. Re-suspend cells in 200uL cold PBS and vortex.
3. In a 15mL conical centrifuge tube (or 12X75mm round-bottom tube) vortex 4mLs ice cold 70% Ethanol and slowly add the 200uL cells in PBS for rapid dispersion. Incubate on ice a minimum of 45 minutes. Overnight fixation (-20⁰C) may be necessary for resolving sub-G₁ peak.
4. Centrifuge (pellet) cells (~1,200 rpm, 10', 4⁰C).
5. Carefully aspirate supernatant and re-suspend cells in **PI Master Mix** (perform a final cell count) at a final cell density of 0.5X10⁶ cells/mL. Incubate at 37⁰C for 30 minutes prior to analysis by flow cytometry. Samples may be stored at 4⁰C, in the dark for up to 3 days.

Reference

Ormerod, M.G., Ed.: Flow Cytometry A Practical Approach Second Edition
1992, Oxford University Press, Inc., New York, NY